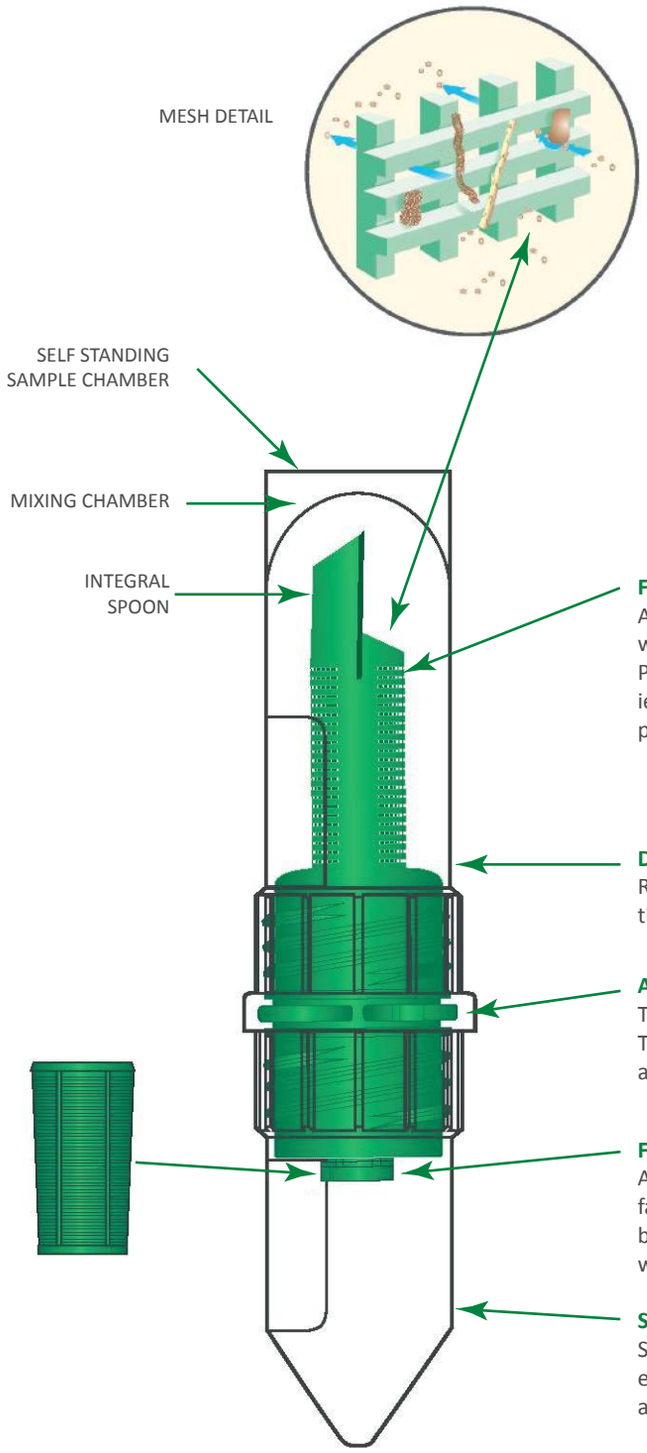
FOR FAECAL CONCENTRATION OF

HELMINTH OVA AND LARVAE / PROTOZOA CYSTS AND OOCYSTS



Filter
A triple stage matrix filtration. Large particles are rejected without obscuring filtration. Recovery rate with Parasep® is comparable to traditional sieve method, ie: Ridley-Allen. The vertical filter enclosed design is patented.

Debris Trap
Rejected particles are trapped to prevent extrusion into the Sedimentation Cone during centrifugation.

Air/Liquid Seal and Safety Lock
The 'seal' prevents the release of biohazardous material. The 'lock' ensures the Mixing Chamber and Filter are removed together for safe disposal.

Fat Dispersion Chamber
A perforated fat dispersion chamber removes the smaller faecal debris and separates the fat content so that it can be efficiently removed from the resulting sediment without the use of ether or ethyl acetate.

Sedimentation Cone
Sediment forms in the base of the cone allowing examination for the presence of helminth eggs or larvae and protozoa cysts or oocysts.

Health and Safety Benefits

- Totally enclosed/sealed process
- Reduced reagent volumes
- No cleaning required
- Single use, no sample contamination
- Ready to use systems available

Performance Benefits

- Optimum sample recovery
- Enhanced sample clarity
- Rapid four step process
- Human resources optimised
- Easy patient identification
- Fits all 50ml centrifuge buckets

PARASITOLOGY

SINGLE USE IN VITRO DIAGNOSTIC DEVICE

APACOR

Midi Parasep® SF

FAECAL PARASITE CONCENTRATOR

Midi Parasep® SF

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Procedure

STEP 1 - SAMPLE PREPARATION

Preserved Samples

Shake or vortex the incoming preserved sample to thoroughly mix.

Transfer either 2ml (**US Gold Standard**) or 3ml (**ARUP JCM**) of the emulsified stool into the Midi Parasep® SF mixing chamber.

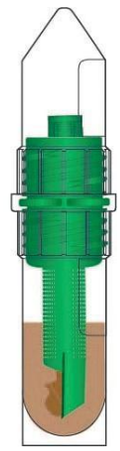
In the event of:

Thick Stool Samples—please add 10 drops of Apacor Triton X solution, then please enclose and vortex/shake to emulsify prior to transferring the sample;

Liquid Stool Samples—please add 4ml instead of 2ml or 3ml to ensure that a sediment is formed after centrifugation is performed.

STEP 2 - EMULSIFICATION

Seal the Midi Parasep® SF by screwing in the filter/sedimentation cone unit.



STEP 3 - CENTRIFUGATION

Invert Midi Parasep® SF and perform centrifugation at:

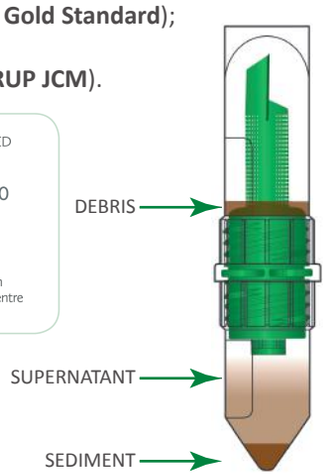
500g for ten minutes (**US Gold Standard**);

400g for two minutes (**ARUP JCM**).

NOTE: TO CALCULATE THE REQUIRED RPM FOR ANY CENTRIFUGE.

$$RPM = \sqrt{\frac{g}{1.12r}} \times 1000$$

RPM - rotor speed in revs/min.
g - centrifugal force (max.1000g)
r - radius, horizontal distance between sedimentation cone tip and spindle centre measured in mm.



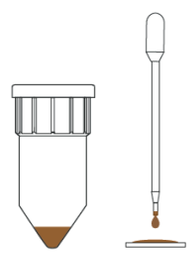
STEP 4 - EXAMINATION

Unscrew and discard the filter and mixing tube.

Decant the supernatant.

Transfer sediment to slide to perform examination where all temporary (Lugol's Iodine) and permanent staining techniques (Trichrome) can be conducted providing a universal fixative is used.

If a universal fixative is used in conjunction with this device, then molecular and EIA tests can be conducted from the sediment.



See label for storage conditions and expiry date. Please adhere to the following guidelines when handling Midi Parasep® SF. To avoid cross contamination the Midi Parasep® SF device should remain closed at all times except when introducing the sample or when retrieving the final concentrated sample for examination.

Midi Parasep® SF is available empty or reagent ready
Please ask for details

Products can be ordered direct from Apacor or from an appointed distributor
Visit our website for all the latest information www.apacor.com or email on: orders@apacor.com

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